

Review

Overview of *Pleurotus* spp., edible fungi with various functional properties

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Abstract

Pleurotus fungi, also known as basidiomycetous fungi, have been a part of human culture for thousands of years. They exhibit anticancer, antitumor, antibacterial, and immunomodulatory effects, having biotechnological, medicinal, and aesthetic applications. They are also versatile, highly resistant to illnesses and pests, and do not require special growing conditions. These properties make them readily marketable, and can be found in supermarkets worldwide, generating multimillion-dollar sale revenues. The global edible mushroom market was valued at USD 5.08 billion in 2021, which is expected to grow to USD 6.43 billion in 2028. China produces about 87% of *Pleurotus* spp. globally; other Asian countries generate 12%, and Europe and America account for approximately 1%. *Pleurotus* spp. have distinct functional characteristics, including high protein content with a proper essential amino acid score pattern, dietary fibre profile, high amounts of vitamins (e.g., B and D) and minerals (e.g., Fe, Zn, Cu, and Se), and low fat. Therefore, *Pleurotus* spp. can provide alternative industrial tools. The present review discusses *Pleurotus* spp. as biotechnological tools for acquiring metabolites of interest, studying them, and analysing bioactive substances that can be used in various fields, including medicine and food.

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Introduction

Mushrooms are the spore-bearing fruiting bodies of macroscopic fungi. Their applications vary, ranging from obtaining metabolites with potential therapeutical properties (Rathee *et al.*, 2012) to the preparation of traditional “Mexican quesadillas” (tortillas with asadero cheese). *Pleurotus* spp., commonly known as oyster mushrooms, are edible mushrooms with gastronomic, nutritional, and medicinal properties (Figure 1). *Pleurotus* spp. are one of the most important commercially farmed edible mushrooms, with a global estimated annual production of 6.46×10^6 tons (Sánchez and Royse, 2017). *Pleurotus* spp. have numerous benefits,

including in human nutrition, animal feed, medicine, pharmacy, chemical industry, biological control, and soil decontamination (Maftoun *et al.*, 2015; Sánchez and Royse, 2017). To date, approximately 70 species of this genus have been reported; however, only a few are commercially viable, such as *P. ostreatus*, *P. florida*, *P. sajor-caju*, and *P. eryngii* (Maftoun *et al.*, 2015). Approximately 99% of *Pleurotus* spp. are produced on the Asian continent, particularly in China. The commercial output of *Pleurotus* spp. from South America is predominantly from Brazil, Mexico, Colombia, Argentina, and Guatemala (Sánchez and Royse, 2017).

Pleurotus spp. cultivation is simple and inexpensive because it does not require sophisticated

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equipment or technology (Figure 2), or other resources (Tesfaw *et al.*, 2015). *Pleurotus* spp. can grow on a variety of lignocellulosic substrates and other agrowastes such as wheat straw, oat, rice, sawdust, cotton waste, banana leaves, and corn stalks. It has a short cultivation cycle, and grows between 25 and 30°C (Bautista *et al.*, 1998; Sánchez, 2004; Sánchez and Royse, 2017).

To date, studies on *Pleurotus* spp. have led to the generation of patents in various fields, including medicines, foods, and cosmetics (WIPO, 2020)

(Table 1). Therefore, research that confirms the biological activities of *Pleurotus* spp., and outlines strategies to obtain specific compounds are crucial. In the present review, we examine the importance of *Pleurotus* spp. as a therapeutic agent, their functional qualities and biomolecules involved, and mechanism of action. The information presented herein will also highlight the economic value of *Pleurotus* spp. in the food business. Finally, we discuss our thoughts on future studies on *Pleurotus* spp.

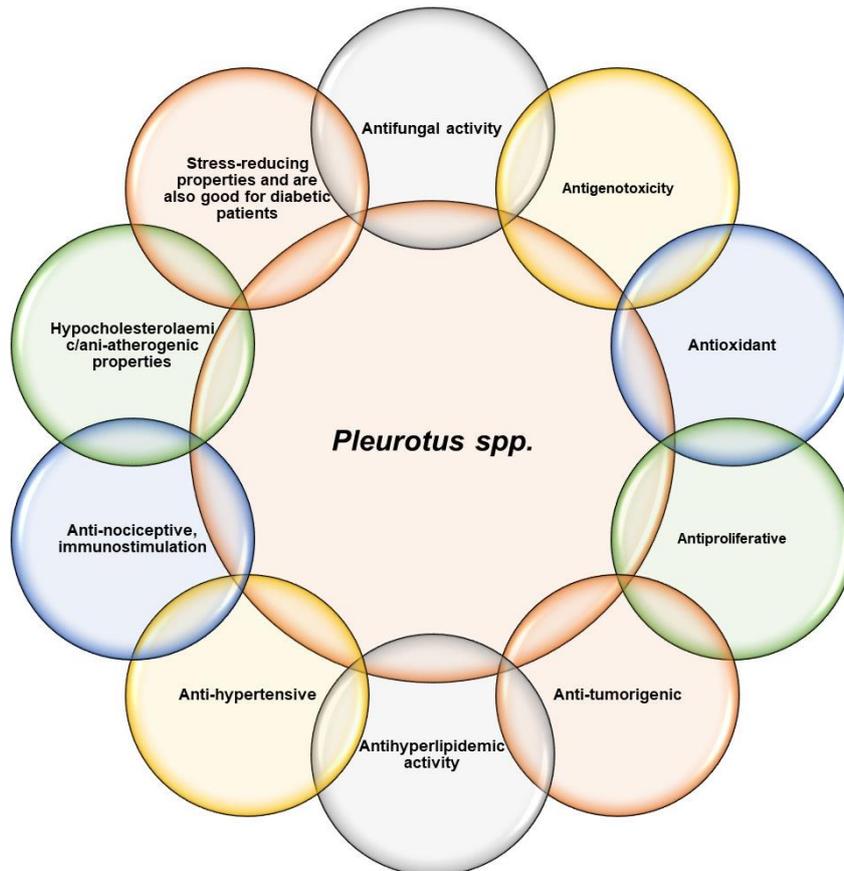


Figure 1. Physiological effects exerted by metabolites produced by *Pleurotus* spp.

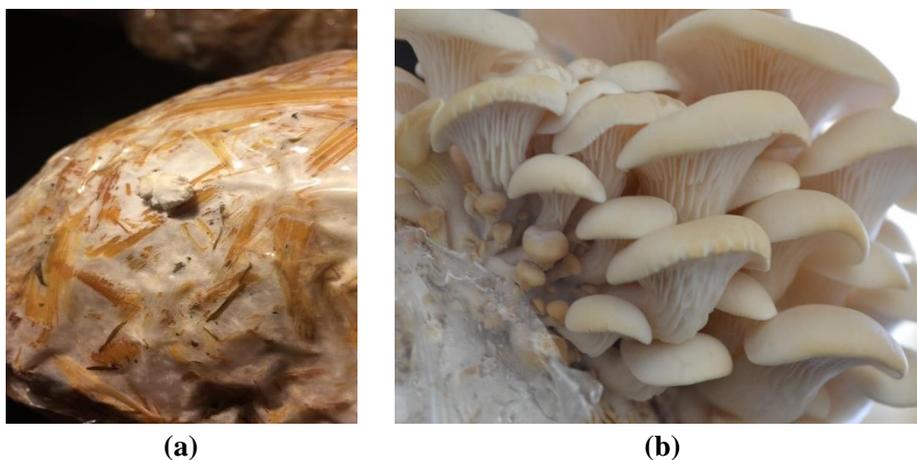


Figure 2. *Pleurotus* spp. production. (a) *Pleurotus* spp. mycelium inoculated in sterile straw as substrate and incubated for approximately 21 days. (b) Fruiting bodies of *Pleurotus* spp. growing on the substrate.

Table 1. Examples of patents generated with *Pleurotus* spp.*

Publication number	Title	Species	Use
109091422	Cosmetics containing <i>P. citrinopileatus</i> extracting solution and preparation method thereof	<i>P. citrinopileatus</i>	Cosmetics
1999169193	Production of trehalose from fungus belonging to genus <i>Pleurotus</i>	<i>P. eryngii</i> (ATCC No.36047), <i>P. cystidiosus</i> (ATCC No. 48751), <i>P. sajor-caju</i> (ATCC No.32078)	Foods, cosmetics, medicines
109293572	Method for extracting ergothioneine and polysaccharides from <i>P. citrinopileatus</i>	<i>P. citrinopileatus</i>	Cosmetics
1995258062	Cosmetics	<i>P. cornucopiae</i>	Cosmetics
10199500292765-	Methyl-5-0-(alpha-d-xylopyranosyl)-d-erythroascorbic acid	<i>P. ostreatus</i>	Therapeutic agents, food additives
WO/2007/132900	Skin moisturizer and therapeutic agent for dermatitis	<i>P. cornucopiae</i>	Therapeutic agent-dermatitis
2001064194	Therapeutic agent of hyperlipemia and therapy of hyperlipemia	<i>P. eryngii</i>	Therapeutic agent-hyperlipemia
2009029786	Pancreatic lipase inhibitor, its production method and therapeutic method	<i>P. eryngii</i>	Therapeutic agent-pancreatic lipase inhibitor

*WIPO (2020).

Nutritional value of Pleurotus

Edible mushrooms are commonly used as functional foods, or as raw materials for functional foods. They are also an excellent food supplement (Kakon *et al.* 2012). In healthcare, they are used to

treat diseases such as infections, diabetes, and hypertension owing to their nutritional value (Figure 3). Some of the most important biomolecules present in *Pleurotus* spp. are described in the following sections.

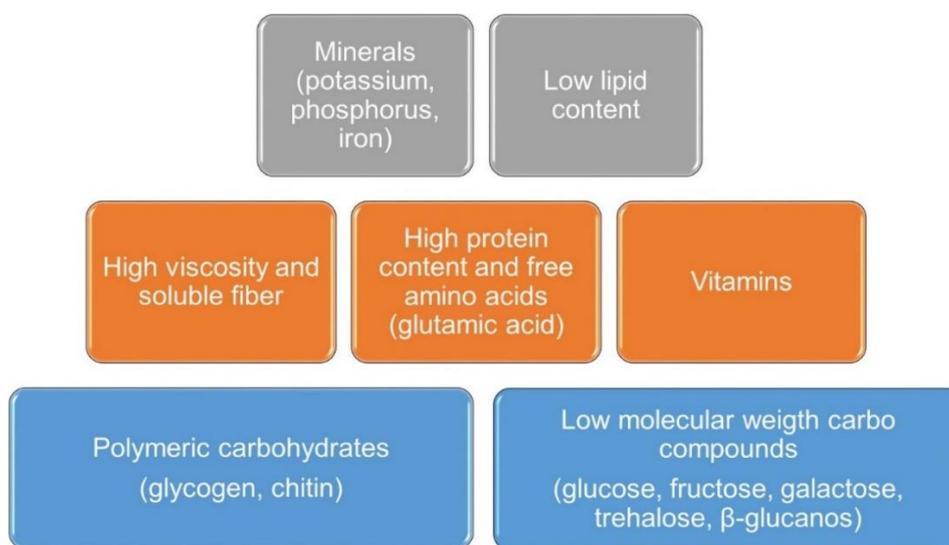


Figure 3. Nutritional components of *Pleurotus* spp.

Proteins, lectins, and amino acids

The protein content of *Pleurotus* spp. is important, and their nutritional value has been recognised for a long time. These fungal proteins have better nutritional value than many plant proteins, with their quality resembling that of animal proteins (Lelley, 1987; da Silva *et al.*, 2012). The protein content of *Pleurotus* spp. (9.29 to 37.4 per 100 g) is higher than that in rice (7.1 to 8.3 g per 100 g) (Juliano, 1985), which is an important staple food (FAO, 2020). The protein content of *Pleurotus* spp. is affected by the species and strain, in addition to the maturation stage, fungal body, collection location, and substrate composition where the fungus was cultivated (Bano and Rajarathnam, 1988; Alam *et al.*, 2008). *Pleurotus* spp. cultivation is essential because agrowastes without apparent use are transformed into biomass with high-quality protein content (Wang *et al.*, 2001; Ancona *et al.*, 2005; Gupta *et al.*, 2013; Fernandes *et al.*, 2015).

Owing to its protein benefits, *Pleurotus* spp. have been used as an alternative to improve the quantity of protein in different foods. For example, *P. sajor-caju* powder (25 - 50%) has been added to chicken and beef burgers. Sensory evaluation showed that consumers accepted hamburgers with different concentrations (25 and 50%) of mushroom powder. In these products, the preference scores for the sensory attributes (aroma, colour, elasticity, juiciness, and flavour) and acceptability were not significantly different from those of the unfortified samples (Wan Rosli *et al.*, 2011; Wan Rosli and Solihah, 2012). In addition, Saiful Bahri and Wan Rosli (2016) investigated the effect of replacing coconut milk powder with *P. sajor-caju* (4 and 20%) on the nutritional composition and sensory acceptability of ready-to-eat pasta in Malaysia. The formulations had high and low protein fat contents. In addition, consumers accepted formulations containing more than 40% mushroom powder (Saiful Bahri and Wan Rosli, 2016). Additionally, replacing wheat flour with a low concentration (5%) of *P. ostreatus* powder-fortified bread did not negatively affect sensory acceptability (Ndung'u *et al.*, 2015). Ng *et al.* (2017) found that 8% supplementation with *P. sajor-caju* powder in cookies resulted in a more desirable aroma, colour, and taste, as compared to cookies without supplementation.

On the other hand, *Pleurotus* contains lectins which are proteins that do not exhibit enzymatic activity. However, lectins can recognise and bind

selectively, specifically, and reversibly to carbohydrates. Lectins are also involved in biological functions such as cell-cell recognition, cell adhesion, and innate immune response against pathogens (Perduca *et al.*, 2020). Furthermore, exhibiting antiproliferative activity against human cancer cells is one of the most critical functions of lectins, thus making them crucial compounds in cancer treatment (Perduca *et al.*, 2020). Lectins obtained from *P. citrinopileatus* have displayed antitumor and antiviral effects (Li *et al.*, 2008). Moreover, a lectin present in *P. ostreatus* (POL) reduced tumour burden in S180 sarcoma and H-22 hepatoma in mice by 88.4 and 75.4%, respectively (Wang *et al.*, 2000). Lectins isolated from *P. florida* reduced arsenic-induced cytotoxicity in rat renal cells by inhibiting apoptosis (Rana *et al.*, 2015).

Pleurotus spp. are suitable for children, adolescents, and adults (FAO, 2013). *Pleurotus* spp. also contain significant percentage (20 - 40%) of free essential amino acids, which are required for a healthy human diet (Maftoun *et al.*, 2015; Grabarczyk *et al.*, 2019). Leucine and glutamic acid are the most abundant essential and non-essential amino acids, respectively (Kayode *et al.*, 2015; Maftoun *et al.*, 2015). Another amino acid of significance within the *Pleurotus* genus is ergothioneine which contains a thiol group. Its biological functions include displaying antioxidant activity (protecting the cells from oxidative/nitrosative stress) and preventing and treating atherosclerosis (Abidin *et al.*, 2017; Izham *et al.*, 2022).

Carbohydrates

Pleurotus spp. contain carbohydrates, particularly polymeric molecules, such as β -glucan, glycogen, chitin, and various low-molecular-weight carbonaceous compounds (*e.g.*, glucose, fructose, galactose, and trehalose). Carbohydrates, along with other components including mannoproteins, galactomannans, celluloses, and polyglucuronic acids, play a key part in the therapeutic effects of mushrooms owing to immune-stimulating glucans (Batbayar *et al.*, 2012; Dalonso *et al.*, 2015; Ng *et al.*, 2017).

β -glucan is one of the main components of the *Pleurotus* spp. dietary fibre (Table 2). This polysaccharide is a soluble fibre with a β -(1 \rightarrow 3) backbone linked to D-glucose with no branches, or varying amounts of β -(1 \rightarrow 6) branches. The glucose chains of β -glucans twist and form a single or triple

helix stabilised by interchain hydrogen bonds (Brown and Gordon, 2005; Rop *et al.*, 2009). The relative molecular weight of β -glucans ranges from tens to thousands of kilodaltons (Rop *et al.*, 2009). This polysaccharide can reduce elevated serum cholesterol levels (Cheung, 2013). Adding 8% *Pleurotus* powder to biscuits increased dietary fibre content from 3.37 to 8.62%, and decreased glycaemic index *in vivo*. This effect was attributed to the fungal fibre which interfered with the starch granules by reducing their size and inducing uneven and spherical shapes, thus resulting in lower susceptibility of the starch to digestive enzymes (Ng *et al.*, 2017). β -glucans possess other beneficial properties, including anti-inflammatory, antitumor, antimicrobial, and antidiabetic activities (Chong *et al.*, 2016). For example, carboxymethylated α -(1 \rightarrow 3)-glucan from *P. citrinopileatus* showed cytotoxic activity in cervical cancer cells (Wiater *et al.*, 2011; 2015), whereas (1 \rightarrow 3)- β -D-glucan and mannogalactan of *P. sajor-caju* had an anti-inflammatory effect (Silveira *et al.*, 2014; 2015). Besides, the PAP polysaccharide

from *P. abalonus* showed antiproliferative activity against human colorectal cancer cells LoVo (Ren *et al.*, 2015).

It is important to note that edible fungi have also been demonstrated to act as prebiotics by stimulating the expansion of the gut microbiota, thereby providing health benefits to the host. Carbohydrates such as chitin, hemicellulose, glucan, mannan, xylan, and galactan are thought to be responsible for this effect (Jayachandran *et al.*, 2017).

Polysaccharides derived from *Pleurotus* spp. can play an essential role in improving the properties of fermented milk products. For example, a polysaccharide derived from *P. eryngii* was added to milk before fermentation, and its effect was examined. The fermented dairy had acceptable firmness and rubberiness values when compared with the controls (Li and Shah, 2015). Additionally, some lactic acid bacteria produced polysaccharides during fermentation, which were necessary for the optimal consistency and texture of the fermented milk products (Zisu and Shah, 2003).

Table 2. Type and quantity of polysaccharides in several species of *Pleurotus*.

Species	Polysaccharide	Dry mass in 100 g	Reference
<i>P. citrinopileatus</i>	total glucans	18.260 g	Sari <i>et al.</i> (2017)
<i>P. eryngii</i>	β -glucans	15.321 g	
<i>P. ostreatus</i> *	β -glucans	4.6 g	Nitschke <i>et al.</i> (2011)
<i>P. pulmonarius</i> *	β -glucans	2.5 g	

*Mycelium

Phenolic compounds

Phenolic compounds are molecules with aromatic rings containing one or more hydroxyl groups. The functions of phenolic compounds include stabilising lipid oxidation and antimicrobial activity (Torres-Martínez *et al.*, 2022). Examples of phenolic compound classes include simple phenolic compounds, phenolic acids, flavonoids, flavones, and phenylpropanoids (Cateni *et al.*, 2022).

Phenolic compounds have been reported in *P. ostreatus*, *P. djamor*, *P. florida*, *P. citrinopileatus*, and *P. giganteus*, in amounts of approximately 700 mg/100 g dry mass (Izham *et al.*, 2022). Flavonoids, mainly catechin, quercetin, and chrysin, are the most abundant phenolic compounds in *Pleurotus* spp. (Mohamed and Fargahaly, 2014). Furthermore, several phenolic acids—such as benzoic acid derivatives (protocatechuic, gallic, vanillic, and syringic acids) and cinnamic acid derivatives (caffeic

and ferulic acids)—have been reported in *P. djamor* (Izham *et al.*, 2022).

Phenolic compounds and secondary metabolites obtained from *P. florida* extracts have antioxidant and antimicrobial activities (Prabu and Kumuthakalavalli, 2016). Flavonoids from *P. djamor* possess similar properties but act as metal chelators, reducing agents, free radical scavengers, and deactivators of singlet oxygen (Sudha *et al.*, 2016).

Vitamins and minerals

Vitamins and minerals exhibit various health benefits in humans. *Pleurotus* spp. are rich in vitamins and minerals but low in fat and calories, thus making it an ideal food for people with diabetes (Kumar *et al.*, 2021).

Vitamins have been reported to exhibit antioxidant activity (Zehiroglu and Ozturk Sarikaya, 2019). *Pleurotus* spp. are particularly rich in thiamine

(vitamin B₁), riboflavin (vitamin B₂), niacin (vitamin B₃), pantothenic acid (vitamin B₅), ascorbic acid (vitamin C), and biotin (vitamin H) (Grabarczyk *et al.*, 2019; Torres-Martínez *et al.*, 2022). Additionally, *Pleurotus* spp. contain tocopherol (vitamin E) and ergosterol. Moreover, *Pleurotus* spp. contain folic acid (vitamin B₉), which decreases the risk of anaemia, diabetes, and high blood pressure (Galappaththi *et al.*, 2021).

Pleurotus spp. contain elements such as potassium, phosphorus, calcium, magnesium, sodium, copper, zinc, iron, manganese, cadmium, lead, and selenium (Khatun *et al.*, 2012; Lavelli *et al.*, 2018; Torres-Martínez *et al.*, 2022). Minerals are essential nutrients involved in different metabolic processes and nerve impulse transmission (Gogavekar *et al.*, 2014). The metal elements found in *Pleurotus* spp. are found in low concentrations; therefore, their consumption would not pose a health risk (Egra *et al.*, 2019).

Lipids

Low levels of lipids are present in edible fungi. In *Pleurotus* spp., it has been reported to range from 0.5 to 20% of fat in dry weight (Grabarczyk *et al.*, 2019; González *et al.*, 2021). These variations depend on the species and the substrate on which they are growing (González-Tijera *et al.*, 2014; Torres-Martínez *et al.*, 2022). Lipids are mono- and polyunsaturated fatty acids (Sande *et al.*, 2019). Linoleic and oleic acids are essential lipids; they are saturated, and the most abundant fatty acids in *P. ostreatus* and *P. levis* (González-Tijera *et al.*, 2014). Linoleic acid (omega-6) is anticarcinogenic, and helps to reduce the risk of cardiovascular diseases and the expression of pro-inflammatory compounds. Besides, it has an essential effect as an inhibitor of the cycle of oxidative stress (COX) (Schneider *et al.*, 2011; Ha *et al.*, 2020). The linoleic and oleic acids also exhibit antioxidant activity (Gnanwa *et al.*, 2021). Another example of a highly abundant saturated fat found in *P. ostreatus* and *P. levis* is palmitic acid which exhibits selective cytotoxicity against human leukemic cells, and induces apoptosis in the MOLT-4 human cell line. It also stimulates glucose incorporation into the adipocytes (Ragasa *et al.*, 2015).

On the other hand, the presence of ergosterol has been found in *P. florida*, *P. djamor*, and *P. eryngii* (Ragasa, 2018). Ergosterol is a precursor of vitamin D₂ produced *via* exposure to UV light, and exhibits

antioxidant activity and prevents cardiovascular diseases (Selvamani *et al.*, 2018; González *et al.*, 2021; Koutrotsios *et al.*, 2022). *P. florida* also contains ergosterol peroxide and cerevisterol (Marquez-Fernandez *et al.*, 2014; Ragasa *et al.*, 2015) which have an inhibitory effect on the NO production, and in the peroxy radical-scavenging activity (Ha *et al.*, 2020).

Molecules and extracts involved in anticancer, hypolipidemic, and anti-obesity activity

Pleurotus spp. are rich in biomolecules which play an essential role in human health. These biomolecules possess a variety of therapeutic properties and have different medicinal qualities, including anticancer, hypolipidemic, and anti-obesity.

Anticancer activity

In recent years, *Pleurotus* spp. have gained importance because of its ability to combat cancer. It has been observed that crude extracts and chemical fractions from *Pleurotus* fruiting bodies or mycelium contain compounds that exhibit anticancer effect toward several types of tumour cells, but this activity depends on the species. Extracts from *P. pulmonarius*, *P. florida*, and *P. djamor* have demonstrated efficacy against several cancer types or cell lines, including human hepatoma HepG2 cells, bladder carcinoma, and ovarian cancer, respectively (Selvi *et al.*, 2011; Xu *et al.*, 2012; Ragasa, 2018).

Polysaccharides, proteins, lectins, vitamins, and selenium from *Pleurotus* spp. have play an important role in the anticancer activity. For instance, according to Cao *et al.* (2015), the polysaccharide POMP2 isolated from *P. ostreatus* mycelium suppressed the proliferation of the BGC-823 human gastric cancer cell line in a concentration-dependent manner, and lowered their ability for invasion. Additionally, an *in vivo* test showed that POMP2 could stop the development of tumours in mice. In this case, the sarcoma 180 sizes were reduced by around 75% by the so-called fraction II polysaccharides from *P. ostreatus* mycelium with doses of 10 and 30 mg/kg (Wisbeck *et al.* 2017). Besides, the extracellular polysaccharides obtained from *P. djamor* showed a tumour inhibition rate of 94% on the sarcoma 180 animal model at a dose of 30 mg/kg (Borges *et al.*, 2014).

Similarly, various techniques are used to extract molecules, which may impact the

effectiveness of the recovered compounds as anticancer agents. Polysaccharides from *P. ostreatus* fruiting bodies showed cytotoxic activities toward Ehrlich ascites carcinoma cell line, suppressing more than 90% of the abnormal cells with a dose of 1000 g/mL (Uddin *et al.*, 2019). Although selenium was considered dangerous to humans in the 1930s, its introduction to *Pleurotus* cultures boosted protein content and antioxidant activity, which is connected to anticancer activities (Khandrika *et al.*, 2009; Kaur *et al.*, 2018).

It has been noted that consumption of *P. ostreatus* supplemented with selenium and zinc during the cultivation process decreases the number of tumour nodules in mice lungs. Besides, an increment in the glutathione peroxidase and superoxide dismutase activities was observed (Yan and Chang, 2012). The ethanolic extract from the mycelium, which lacking selenium in the culture medium, had a poor cytotoxic effect on LS174 and He-La cells (Milovanovic *et al.*, 2014).

On the other hand, selenium polysaccharide SPMP-2a from *P. geesteranus* decreased the amount of swollen and vacuolar mitochondria in the H₂O₂-treated cells as compared to the controls. Besides, this polysaccharide reduced the amount of reactive oxygen species, and improved the catalase and superoxide dismutase activities (Sun *et al.*, 2017). The activation of apoptotic mechanisms, which are produced by the generation of reactive species (Wang *et al.*, 2014; Yang *et al.*, 2018) or due to the up-down regulation of numerous proteins are thought to be the causes of *Pleurotus* anticancer activities. For instance, in a concentration-dependent manner human colorectal adenocarcinoma cell SW480 viability was decreased by *P. ostreatus* protein extract. The production of reactive oxygen species, glutathione depletion, and a loss in mitochondrial transmembrane potential are the apoptotic mechanisms that result in the proteins present in the *P. ostreatus* extract on SW480 cells (Wu *et al.*, 2011); however, the type of proteins present in this extract is unknown.

Interestingly, human breast (MCF-7) and colon cancer (HT-29) cell proliferation is inhibited by using *P. ostreatus* methanolic extracts. The effect might have been due to an up-regulation of p53 and p21 that arrested the G0/G1 cell cycle (Jedinak and Silva, 2008). Likewise, the water-soluble proteoglycan from *P. ostreatus* arrested the Sarcoma-180-bearing mice model cells in the pre-G0/G1 phase (Sarangi *et*

al., 2006). Besides, it has been detected that extracts from *P. highking* induce apoptosis in breast cancer cells by altering the balance of proapoptotic and antiapoptotic genes (Haque and Islam, 2019). Also, the viability of HT-29 human colon cancer cells was reduced by 60.1 and 59.3%, respectively, using methanolic extracts from the fruiting bodies of *P. ostreatus* and *P. salmoneostramineus* (Kim *et al.*, 2009).

Based on previous findings, *Pleurotus* spp. exhibited anticancer characteristics. However, there may be variations in the anticancer activities and efficacies, depending on the species and the variety of the extracted compounds.

Hypolipidemic activity

It has been reported that the use of 5% of *P. ostreatus* fruiting bodies in the diet of hypercholesterolemic rats decreased the plasma triglyceride, cholesterol, and total lipid and phospholipid levels (Alam *et al.*, 2011). Likewise, a hypolipidemic effect was observed by using the aqueous extract or pellets of *P. eryngii* in the diet of laboratory animals (Mizutani *et al.*, 2010; Choi *et al.*, 2017). It was found that polysaccharides from *P. eryngii* mycelium extracted by alkaline or acid methods had hypolipidemic activity (Ren *et al.*, 2017; Xu *et al.*, 2017). Also, exopolysaccharides from *P. geesteranus* were responsible for decreasing in cholesterol and triacylglycerol concentrations (Dubin *et al.*, 2013). Huang *et al.* (2014) suggested that β -glucans and fibre contained in the exopolysaccharides of *P. tuber-regium* might be responsible for the hypolipidemic activity. However, the mechanism for this action is unknown. The same authors pointed out that polysaccharides may act as agonists of a transcriptional factor (PPAR- α) that regulates genes involved in lipid metabolism.

Chrysin, a flavonoid with antioxidant activity from *Pleurotus* has a hypolipidemic effect in rats. This phenomenon can be due to the reactivation of a mechanism involving lipolytic enzymes (Anandhi *et al.*, 2013). Interestingly, extracts from *P. citrinopileatus* containing nicotinic acid and ergosterol reduced triglycerides and cholesterol in rats (Schneider *et al.*, 2011; Zeb *et al.*, 2013).

Anti-obesity activity

Obesity has become one of the most critical health problems worldwide, requiring an urgent need for efficient control. In this regard, *Pleurotus* spp. are

rich in bioactive molecules that can contribute to reaching this goal. For example, in one study, the anti-obesity and lipid-lowering effect of *P. citrinopileatus* water extract (PWE) was evaluated using a series of biochemical assays in randomised C57BL/6J mice with high-fat diet-induced obesity (DIO). Results showed that PWE significantly reduced the mice's weight gain, fat accumulation, and food intake. PWE also decreased serum triglycerides, cholesterol, and low-density lipoprotein, and improved glucose tolerance in high-fat-fed mice (Sheng *et al.*, 2019). The role of *P. ostreatus* in reducing obesity, maintaining glucose homeostasis, and modulating the gut microbiota has been demonstrated in mice (Hu *et al.*, 2022). Also, it has been detected that polysaccharides from *P. eryngii* had a positive effect in reducing obesity in mice. Data showed that *P. eryngii* polysaccharides had anti-obesity and LDL cholesterol-lowering effects in obese mice, probably through an increment in the excretion of bile acids and lipids, and its impact in altering the microbiota (Nakahara *et al.*, 2020).

Other biological activities of Pleurotus spp.

Extracts from various *Pleurotus* spp. have been found to have a variety of actions, including anti-inflammatory, anticancer, and proinflammatory properties. The extract components induce the immune response, signal transduction, and phagocytic activities. For example, Table 3 shows a list of crude extracts, their active molecules, and physiological effects (Paulík *et al.*, 1996; Nozaki *et al.*, 2008; Jedinak *et al.*, 2011; Xu *et al.*, 2014; Hu *et al.*, 2018; Ma *et al.*, 2020; Llauroadó *et al.*, 2021).

Mutants and overexpression of genes

Different molecular methods have enabled the transformation of *Pleurotus*. The generation of recombinant strains has enhanced our knowledge about the function of other genes, and the possibility of increasing the nutritional properties, fruit body quality, pathogen resistance, and biotic-abiotic stresses, thus increasing productivity, enhancing shelf life, and producing sporeless/low-sporing strains. These improvements have been accomplished using selection techniques, hybridisation, mutation breeding, molecular breeding, genetic transformation, and genome editing techniques (Ravishankar *et al.*, 2006; Barh *et al.*, 2019).

Different protocols have been developed to transform *Pleurotus* spp., such as polyethylene

glycol/CaCl₂, electroporation, particle bombardment, and *Agrobacterium*-mediated transformation (Honda *et al.*, 2000; Sunagawa and Magae, 2002; Sharma and Kuhad, 2010; Barh *et al.*, 2019). Recently, CRISPR/Cas9 has been used for efficient genome editing in *P. ostreatus*. This powerful tool can contribute to the molecular breeding of non-genetically modified strains that can contribute to the acceptance in different countries (Boontawon *et al.*, 2021).

For instance, knockout and overexpression of a carbon catabolite repressor (*cre1*) in *P. ostreatus* PC9 generated mutants that showed an increase or decrease in the cellulolytic activity. However, overexpression of *cre1* did not produce a higher glucose yield relative to the wild type (Yoav *et al.*, 2018).

In contrast, the overexpression of the enzyme manganese peroxidase *pomnp6* and versatile peroxidase *povp3* in *P. ostreatus* led to higher lignin degradation in cotton stalks. These data suggested that *pomnp6* and *povp3* might help develop sustainable energy (Wang *et al.*, 2019). Likewise, the protein kinase A catalytic subunit (*PKAc*) is essential for inducing the wood degradation system in *P. ostreatus*. Overexpression of *PKAc* in *P. ostreatus* resulted in faster degradation of beechwood lignin (Toyokawa *et al.*, 2016). *Pleurotus* spp. use the ligninolytic system to transform agrowastes into products that can be used for their growth, thus producing valuable food products (Cohen *et al.*, 2002). Another gene overexpressed in *P. ostreatus* is the methionine sulfoxide reductase A which is encoded for the *PoMsrA* enzyme that reverses the oxidation of methionine sulfoxide (oxidised methionine). The overexpression of *PoMsrA* generated tolerance at high temperatures, osmotic pressure, and oxidative stress, thus suggesting its role in cellular protection against oxidative stress (Yin *et al.*, 2015). The production of β -glucans in *P. ostreatus* was improved by replacing the promoter of the β -1,3-glucan synthase gene (*GLS*) with the promoter of the glyceraldehyde-3-phosphate dehydrogenase gene from *Aspergillus nidulans*. This transformation led to the overexpression of *GLS*, and consequently, the overproduction of β -glucans (Chai *et al.*, 2013).

The role that *Pofst3* plays during the development of *P. ostreatus* was assessed through overexpression and antisense silencing *via Agrobacterium*-mediated transformation. The *Pofst3*-

Table 3. Example of crude extracts, active molecules obtained from *Pleurotus*, and their physiological effect.

Biological activity	Species	Type of extract	Possible molecule responsible	Involved molecule and its effect	Reference
Anti-inflammatory activity	<i>P. ostreatus</i>	Water extract	ND	Transcription activity ↓ AP-1, ↓ NF-κB Production ↓ TNF-α, ↓ IL-6, ↓ IL-12, ↓ PGE2, ↓ NO, ↓ IFN-γ, ↓ IL-2	Jedinak et al. (2011)
Immune response	<i>P. eryngii</i>	Saponified extract	Acidic glycosphingolipid	Secretion ↑ IFN-γ, ↑ IL-4	Nozaki et al. (2008)
Antitumor activity	<i>P. ferulae</i> , wild and cultivated	Protein purified from fresh fruiting bodies	PEP 1b Methyl linoleate, hexadecanoic acid methyl ester, and other compounds	Production ↑ NO, ↑ IL-1β, ↑ IL-6, ↑ TNF-α ↑ apoptosis partially mediated by reactive oxygen species	Hu et al. (2018) Wang et al. (2014); Yang et al. (2018)
Proinflammatory activity	<i>P. ostreatus</i>	Aqueous	ND	↑ NO, ↑ iNOS expression, ↑ TNF-α, ↑ IL-6 ↑ expression Mif, ↑ expression Rap 1b, MAPK pathway ↑ Gpnmb, ↑ Sod1, ↑ C5ar1, ↑ Prdx2	Llauradó et al. (2021)
Signal transduction	<i>P. eryngii</i>	Protein isolate	PEP 1b	Nitric oxide biosynthetic process NF-κB signalling pathway ↑ Cox2, ↑ Hsp90aa1 ↑ Pyk2, ↑ Itgb2 ↑ expression Sqstm1, ↑ Cox2, ↑ Itgb2	Ma et al. (2020)
Hemagglutination	<i>P. ferulae</i>	Aqueous and 75% saturated (NH ₄) ₂ SO ₄	Lectin	ND	Xu et al. (2014)
Phagocytic activity	<i>P. ostreatus</i>	Purification of the soluble form	Glucan	ND	Paulík et al. (1996)

ND: Not determined.

overexpressing strains showed fewer primordia and larger fruiting bodies; however, no statistically significant differences were observed between these strains and the wild-type strain. Nevertheless, the strains with silenced *Pofst3* developed more primordia, and showed smaller fruiting bodies. Therefore, it was observed that *Pofst3* was involved in inhibiting primordial cluster formation and regulating the development of fruiting bodies in *P. ostreatus*. The efficient production of this edible mushroom may benefit from improvements in its development, thereby increasing its possible biotechnological applications (Qi *et al.*, 2019). Furthermore, laccase production by *P. eryngii* var. *ferulae* was improved *via* overexpression of *sspoxa3a/b*, which resulted in small fungal pellets and thin mycelial walls, thus facilitating laccase secretion, and increasing extracellular laccase activity. The increase in laccase quantity may interest the food and chemical industries because laccase has been used for its catalytic properties (Zhang *et al.*, 2021).

Mutations could improve certain properties in *Pleurotus* spp. (Barh *et al.*, 2019). For example, Djajanegara and Harsoyo (2008) generated a mutant PO-4 derived from *P. florida* using gamma radiation which showed a higher antioxidant content relative to the control. Also, sporeless or low-sporing mutants of *P. sajor-caju* was generated after 75 min of UV exposure (Ravishankar *et al.*, 2006). Chemical mutagenesis is another tool used for generating genetic diversity. For instance, NQ2A-12, a new variety of *P. eryngii* with improved medicinal qualities, has been developed; in NQ2A-12, the expression of Pin1 was increased. Pin1 is a peptidyl-prolyl *cis-trans* isomerase that represses Alzheimer's disease (Jeong *et al.*, 2017). Random mutagenesis is another technique used to generate *Pleurotus* spp. mutants. For example, a wild strain of *P. ostreatus* was exposed to UV radiation (1.2×10^2 J/m²/S) for incubation periods from 0 to 45 min, and 0.2% of ethidium bromide treatment for exposure periods from 0 to 10 min to enhance the production of manganese peroxidase (MnP). The double-mutated stable strain had an increased MnP activity (368.18 U/L) as compared to the wild strain (86.8 U/L). The aim of this study was to optimise MnP production, as this enzyme is helpful for the biodegradation of textile azo dyes (Arunkumar and Sheik Abdulla, 2014).

Economic value and production

Mushrooms of economic value can be classified differently, mainly as edibles, cosmetics, pharmaceuticals, and wilds (Royse *et al.*, 2017; Grand View Research, 2023). Worldwide, approximately 100 million tons of agrowastes are produced annually, which can cause environmental contamination if they are burned in the open air. Global warming is a problem worldwide, and participating countries in COP26 must contribute to limiting this increment to 1.5°C during the following 20 years (UKCOP, 2021). Fortunately, agrowastes (*e.g.*, corn stalks and leaves) can be used to cultivate edible mushrooms, and using that agricultural biomass in fungal production can contribute to lowering greenhouse gas emissions (Bumanlag *et al.*, 2018). China cultivates the most edible mushrooms worldwide (Niazi and Ghafoor, 2021). The primary cultivated edible fungi and the most consumable in the world are *Lentinula edodes* (shitake), *Pleurotus* spp. (oyster mushrooms), *Auricularia heimuer* (black wood ear), *Agaricus bisporus* (button mushroom), and *Flammulina filiformis* (enokitake) (Li and Xu, 2022; Fortune Business Insights, 2023).

Pleurotus spp. play a significant role in the global mushroom business, accounting for 19 to 25% of the total output (Royse *et al.*, 2017; Raman *et al.*, 2021). China generates approximately 87% of all *Pleurotus* spp., with the remaining 12% originating from other Asian countries (Japan, South Korea, Taiwan, Thailand, Vietnam, and India). Only 1% is grown in Europe and North-South America (Royse *et al.*, 2017). Some of the most cultivated *Pleurotus* spp. are *P. ostreatus*, *P. sajor-caju*, *P. eous*, *P. florida*, and *P. sajor-caju* (Raman *et al.*, 2021). Cultivation of other species such as *P. djamor* utilising agrowastes must also be considered (Bumanlag *et al.*, 2018). On the other hand, the global edible mushroom market (cosmetics, pharmaceuticals, and foods) was valued at approximately USD 50 billion in 2021, respectively, and it is expected to grow annually by 9.7% from 2022 to 2030 (Grand View Research, 2023). In this regard, in 2021, the global *P. ostreatus* market size was evaluated at USD 5.08 billion, with an annual growth of 3.4%, and expected to reach USD 6.43 billion in 2028 (Business Research Insights, 2022).

In South America, the mushroom cultivation market is concentrated in the production of button

mushroom, shitake mushroom, oyster mushroom, and others (nametake, maitake, enokitake, mane mushroom, straw mushroom, shimeji) (Market Data Forecast, 2022), with a global market of USD 4250 million in 2021, and estimated annual growth of 4.8%. Mexico, Brazil, Argentina, Chile, and Colombia are the largest producers of edible mushrooms, and Mexico is the leading producer (EMR, 2023), with around 60% of South-American production (Cano-Estrada and Romero-Bautista, 2016).

These data show the economic significance of *Pleurotus* spp. and other edible mushrooms. In this regard, the yield will probably rise in the following years, not only because these fungi are tasty, but also because of their functional features, thus making them desirable to those looking for natural, minimally processed, and wholesome foods.

Conclusion

As a consequence of different factors such as functional properties, nutritional compositions (carbohydrates, proteins, vitamins, and fibres), and green technology facilities, the consumption of mushrooms, specifically *Pleurotus* spp., has increased. China is a leading mushroom grower, and the production in North and South America is still low as compared to China. Therefore, given the global relevance of *Pleurotus*, farmers and producers should strategically focus on commercial mushroom cultivation, which would be an essential source of income. Furthermore, because fungal manufacturing uses lignocellulosic waste, this activity aids in reducing agrowastes, which are often burned in the field, thus causing air pollution.

While numerous *Pleurotus* patents have been recorded, it is possible to produce new ones, mainly if they are focused on discovering novel metabolites for use in various industries, such as therapeutics, foods, and cosmetics. Even though *P. ostreatus* is the most well-studied *Pleurotus* species; it is vital to concentrate efforts on lesser-studied species, which may contain metabolites with unique functions. Also, several extraction methods have been documented; it is critical to continue researching novel methods to improve the presence of specific activity. Likewise, because there is limited research in this field, it is necessary to expand the development of mutant and transformant lines that overexpress genes of

biotechnological interest that may be employed in diverse domains.

It will also be essential to cultivate edible *Pleurotus* spp. of different colours to satisfy different tastes, and promote their consumption through well-targeted marketing, highlighting their fantastic properties. Besides, different recipes that allow their consumption in different ways, and the development of new products that use flour obtained from these mushrooms (e.g., snacks for children) which contain high protein content should be undertaken. Furthermore, it will be important to create culture methods for wild-type species that allow for rapid commercialisation, and measures to extend their shelf life for commercial purposes should also be investigated most likely by technical or physical process improvements.

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References

- Abidin, M., Abdullah, N. and Abidin, N. 2017. Therapeutic properties of *Pleurotus* species (oyster mushrooms) for atherosclerosis: A review. *International Journal of Food Properties* 20(6): 1251-1261.
- Alam, N., Amin, R., Khan, A., Ara, I., Shim, M. J., Lee, M. W. and Lee, T. S. 2008. Nutritional analysis of cultivated mushrooms in Bangladesh - *Pleurotus ostreatus*, *Pleurotus sajor-caju*, *Pleurotus florida* and *Calocybe indica*. *Mycobiology* 36(4): 228-232.
- Alam, N., Yoon, K. N., Lee, T. S. and Lee, U. Y. 2011. Hypolipidemic activities of dietary *Pleurotus ostreatus* in hypercholesterolemic rats. *Mycobiology* 39(1): 45-51.
- Anandhi, R., Annadurai, T., Anitha, T. S., Muralidharan, A. R., Najmunnisha, K., Nachiappan, V., ... and Geraldine, P. 2013. Antihypercholesterolemic and antioxidative effects of an extract of the oyster mushroom, *Pleurotus ostreatus*, and its major constituent, chrysin, in Triton WR-1339-induced hypercholesterolemic rats. *Journal of Physiology Biochemistry* 69(2): 313-323.

- Ancona, L., Sandoval, C. A., Belmar, R. and Capetillo, C. M. 2005. Effect of substrate and harvest on the amino acid profile of oyster mushroom (*Pleurotus ostreatus*). *Journal of Food Composition and Analysis* 18(5): 447-450.
- Arunkumar, M. and Sheik Abdulla, S. H. 2014. Hyper-production of manganese peroxidase by mutant *Pleurotus ostreatus* MTCC 142 and its applications in biodegradation of textile azo dyes. *Desalination and Water Treatment* 56(2): 1-12.
- Bano, Z. and Rajarathnam, S. 1988. *Pleurotus* mushrooms. Part II. Chemical composition, nutritional value, post-harvest physiology, preservation, and role as human food. *Critical Reviews in Food Science and Nutrition* 27(2): 87-158.
- Barh, A., Sharma, V. P., Annepu, S. K., Kamal, S., Sharma, S. and Bhatt, P. 2019. Genetic improvement in *Pleurotus* (oyster mushroom): A review. *3 Biotech* 9(9): 322.
- Batbayar, S., Lee, D. H. and Kim, H. W. 2012. Immunomodulation of fungal β -glucan in host defense signaling by dectin-1. *Biomolecules and Therapeutics* 20(5): 433-445.
- Bautista, M., Alanis, M. G., González, E. and García, C. L. 1998. Chemical composition of three Mexican strains of mushrooms (*Pleurotus ostratus*). *Archivos Latinoamericanos de Nutrición* 48(4): 359-363.
- Boontawon, T., Nakazawa, T., Inoue, C., Osakabe, K., Kawauchi, M., Sakamoto, M. and Honda, Y. 2021. Efficient genome editing with CRISPR/Cas9 in *Pleurotus ostreatus*. *AMB Express* 11(1): 1-11.
- Borges, G. M., De Barba, F. F., Schiebelbein, A. P., Pereira, B. P., Chaves, M. B., Silveira, M. L., ... and Wisbeck, E. 2014. Extracellular polysaccharide production by a strain of *Pleurotus djamor* isolated in the south of Brazil and antitumor activity on Sarcoma 180. *Brazilian Journal Microbiology* 44(4): 1059-1065.
- Brown, G. D. and Gordon, S. 2005. Immune recognition of fungal β -glucans. *Cellular Microbiology* 7(4): 471-479.
- Bumanlag, C. P. B., Kalaw, S. P., Dulay R. M. R. and Reyes, R. G. 2018. Optimum conditions for mycelial growth and basidiocarp production of *Pleurotus djamor* on corn based media. *International Journal of Biology, Pharmacy and Allied Sciences* 7(4): 558-575.
- Business Research Insights. 2022. *Pleurotus ostreatus* market size, share, growth, and industry analysis, by type (grey oyster mushroom and white oyster mushroom), by application (edible and medicinal) regional forecast to 2028. Retrieved on February 12, 2023 from Business Research Insights website: <https://www.businessresearchinsights.com/market-reports/pleurotus-ostreatus-market-100739#:~:text=The%20global%20pleurotus%20ostreatus%20market%20size%20was%20USD%205.080%20billion%20in%202021>
- Cano-Estrada, A. and Romero-Bautista, L. 2016. Economic, nutritional and medicinal value of edible wild mushrooms. *Revista Chilena de Nutrición* 43(1): 75-80.
- Cao, X. Y., Liu, J. L., Yang, W., Hou, X. and Li, Q. J. 2015. Antitumor activity of polysaccharide extracted from *Pleurotus ostreatus* mycelia against gastric cancer *in vitro* and *in vivo*. *Molecular Medicine Reports* 12(2): 2383-2389.
- Cateni, F., Gargano, M. L., Procida, G., Venturella, G., Cirlincione, F. and Ferraro, V. 2022. Mycochemicals in wild and cultivated mushrooms: Nutrition and health. *Phytochemistry Reviews* 21: 339-383.
- Chai, R., Qiu, C., Liu, D., Qi, Y., Gao, Y., Shen, J. and Qiu, L. 2013. β -glucan synthase gene overexpression and β -glucans overproduction in *Pleurotus ostreatus* using promoter swapping. *PLoS One* 8: e61693.
- Cheung, P. 2013. Mini-review on edible mushrooms as source of dietary fiber: Preparation and health benefits. *Food Science and Human Wellness* 2(3-4): 162-166.
- Choi, J. H., Kim, D. W., Kim, S. and Kim, S. J. 2017. *In vitro* antioxidant and *in vivo* hypolipidemic effects of the king oyster culinary-medicinal mushroom, *Pleurotus eryngii* var. *ferulae* DDL01 (Agaricomycetes), in rats with high-fat diet-induced fatty liver and hyperlipidemia. *International Journal of Medicinal Mushrooms* 19(2): 107-119.
- Chong, V., Al-Azad, S. and Shapawi, R. 2016. Comparison of two edible mushroom extract as aquaculture feed additive to enhance immune response of Asian seabass. *Transactions on Science and Technology* 3(2-2): 427-432.

- Cohen, R., Persky, L. and Hadar, Y. 2002. Biotechnological applications and potential of wood-degrading mushrooms of the genus *Pleurotus*. *Applied Microbiology and Biotechnology* 58(5): 582–594.
- da Silva, M. C. S., Naozuka, J., da Luz, J. M., de Assunção, L. S., Oliveira, P. V., Vanetti, M. C. D., ... and Kasuya, M. C. M. 2012. Enrichment of *Pleurotus ostreatus* mushrooms with selenium in coffee husks. *Food Chemistry* 131(2): 558-563.
- Dalonso, N., Goldman, G. H. and Gern, R. M. 2015. β -(1 \rightarrow 3), (1 \rightarrow 6)-glucans: Medicinal activities, characterization, biosynthesis and new horizons. *Applied Microbiology and Biotechnology* 99(19): 7893-7906.
- Djajanegara, I. and Harsoyo. 2008. White oyster mushroom (*Pleurotus florida*) mutant with altered antioxidant contents. *BIOTROPIA* 15(1): 65-73.
- Doubin, M., Yuping, M., Lujing, G., Aijin, Z., Jianqiang, Z. and Chuping, X. 2013. Fermentation characteristics in stirred-tank reactor of exopolysaccharides with hypolipidemic activity produced by *Pleurotus geesteranus* 5[#]. *Anais da Academia Brasileira de Ciências* 85(4): 1473-1481.
- Egra S., Kusuma, I. W., Arung, E. T. and Kuspradini, H. 2019. The potential of white-oyster mushroom (*Pleurotus ostreatus*) as antimicrobial and natural antioxidant. *Biofarmasi Journal of Natural Product Biochemistry* 17(1): 17-23.
- Expert Market Research (EMR). 2023. Latin American mushroom market – By type (button, Shiitake, oyster, others); by form (fresh, frozen, dried, canned, other); by distribution channel (supermarkets and hypermarkets, grocery stores, online, others); by region (Brazil, Mexico, Argentina, Chile, others); market dynamics, price analysis, competitive landscape. Retrieved on February 13, 2023 from EMR website: <https://www.informesdeexpertos.com/informes/mercado-latinoamericano-de-champinones>
- Fernandes, Â., Barros, L., Martins, A., Herbert, P. and Ferreira, I. C. 2015. Nutritional characterisation of *Pleurotus ostreatus* (Jacq. ex Fr.) P. Kumm. produced using paper scraps as substrate. *Food Chemistry* 169: 396-400.
- Food and Agriculture Organisation (FAO). 2013. Dietary protein quality evaluation in human nutrition. Retrieved on July 5, 2020 from FAO Website: <http://www.fao.org/documents/card/en/c/ab5c9fca-dd15-58e0-93a8-d71e028c8282/>
- Food and Agriculture Organisation (FAO). 2020. FAOSTAT data on food and agriculture. Retrieved on October 1, 2020 from FAOSTAT Website: <https://www.fao.org/faostat/es/#home>
- Fortune Business Insights. 2023. The global mushroom market is projected to grow from 15.25 million tonnes in 2021 to 24.05 million tonnes in 2028 at a CAGR of 6.74% in forecast period. Retrieved on February 12, 2023 from Fortune Business Insights website: <https://www.fortunebusinessinsights.com/industry-reports/mushroom-market-100197>
- Galappaththi, M. C. A., Dauner, L., Madawala, S. and Karunaratna, S. C. 2021. Nutritional and medicinal benefits of oyster (*Pleurotus*) mushrooms: A review. *Fungal Biotech Journal* 1(2): 65-87.
- Gnanwa, M. J., Yorou, N. and Kouamé, L. 2021. Assessment of minerals, vitamins, amino and fatty acids components of *Pleurotus ostreatus* mushrooms cultivated and sold in the village of M'Badon (Abidjan, Côte d'Ivoire). *International Journal of Current Microbiology and Applied Sciences* 10: 276-283.
- Gogavekar, S. S, Rokade, S. A., Ranveer, R. C., Ghosh, J. S., Kalyani, D. C. and Sahoo, A. K. 2014. Important nutritional constituents, flavour components, antioxidant and antibacterial properties of *Pleurotus sajor-caju*. *Journal of Food Science Technology* 51(8): 1483-1491.
- González, A., Nobre, C., Simões, L. S., Cruz, M., Loredó, A., Rodríguez-Jasso, R. M., ... and Belmares, R. 2021. Evaluation of functional and nutritional potential of a protein concentrate from *Pleurotus ostreatus* mushroom. *Food Chemistry* 346: 128884.
- González-Tijera, M., Márquez-Fernández, O., Mendoza-López, M. R., Mata, G. and Trigos, Á. 2014. A comparison of fatty acid content in three species of the genus *Pleurotus*. *Revista Mexicana de Micología* 39: 41-45.
- Grabarczyk, M., Mączka, W., Wińska, K. and

- Ukłańska-Pusz, C. 2019. Mushrooms of the *Pleurotus* genus – Properties and application. *Biotechnology and Food Science* 83(1): 13-30.
- Grand View Research. 2023. Mushroom market size, share and trends analysis report by product (button, shiitake, oyster), by form, by distribution channel, by application (food, pharmaceuticals, cosmetics), by region, and segment forecasts, 2022 - 2030. Retrieved on February 11, 2023 from Grand View Research Website:
<https://www.grandviewresearch.com/industry-analysis/mushroom-market>
- Gupta, A., Sharma, S., Saha, S. and Walia, S. 2013. Yield and nutritional content of *Pleurotus sajor caju* on wheat straw supplemented with raw and detoxified mahua cake. *Food Chemistry* 141(4): 4231-4239.
- Ha, J. W., Kim, J., Kim, H., Jang, W. and Kim, K. H. 2020. Mushrooms: An important source of natural bioactive compounds. *Natural Product Sciences* 26(2): 118-131.
- Haque, A. and Islam, A. U. 2019. *Pleurotus highking* mushroom induces apoptosis by altering the balance of proapoptotic and antiapoptotic genes in breast cancer cells and inhibits tumor sphere formation. *Medicina* 55(11): 716.
- Honda, Y., Matsuyama, T., Irie, T., Watanabe, T. and Kuwahara, M. 2000. Carboxin resistance transformation of the homobasidiomycete fungus *Pleurotus ostreatus*. *Current Genetics* 37: 209-212.
- Hu, Q., Du, H., Ma, G., Pei, F., Ma, N., Yuan, B., ... and Yang, W. 2018. Purification, identification, and functional characterization of an immunomodulatory protein from *Pleurotus eryngii*. *Food and Function* 9(7): 3764-37675.
- Hu, Y., Xu, J., Sheng, Y., Liu, J., Li, H., Guo, M., ... and He, X. 2022. *Pleurotus ostreatus* ameliorates obesity by modulating the gut microbiota in obese mice induced by high-fat diet. *Nutrients* 14(9): 1868.
- Huang, H. Y., Korivi, M., Yang, H. T., Huang, C. C., Chaing, Y. Y. and Tsai, Y. C. 2014. Effect of *Pleurotus tuber-regium* polysaccharides supplementation on the progression of diabetes complications in obese-diabetic rats. *The Chinese Journal of Physiology* 57(4):198-208.
- Izham, I., Avin, F. and Raseetha, S. 2022. Systematic review: Heat treatments on phenolic content, antioxidant activity, and sensory quality of Malaysian mushroom: Oyster (*Pleurotus* spp.) and black jelly (*Auricularia* spp.). *Frontiers in Sustainable Food Systems* 6: 882939.
- Jayachandran, M., Xiao, J. and Xu, B. 2017. A critical review on health promoting benefits of edible mushrooms through gut microbiota. *International Journal of Molecular Sciences* 18(9): 1934.
- Jedinak, A. and Sliva, D. 2008. *Pleurotus ostreatus* inhibits proliferation of human breast and colon cancer cells through p53-dependent as well as p53-independent pathway. *International Journal of Oncology* 33(6): 1307-1313.
- Jedinak, A., Dudhgaonkar, S., Wu, Q., Simon, J. and Sliva, D. 2011. Anti-inflammatory activity of edible oyster mushroom is mediated through the inhibition of NF- κ B and AP-1 signaling. *Nutrition Journal* 10(52): 1-10.
- Jeong, Y., Jung, M., Kim, M. J. and Hwang, C. H. 2017. A 4-nitroquinoleneoxide-induced *Pleurotus eryngii* mutant variety increases Pin1 expression in rat brain. *Journal of Medicinal Food* 20(1): 65-70.
- Juliano, B. O. 1985. Rice: Chemistry and technology. 2nd ed. United States: American Association of Cereal Chemists (AACC).
- Kakon, A. J., Choudhury, B. K. and Saha, S. 2012. Mushroom is an ideal food supplement. *Journal of Dhaka National Medical College and Hospital* 18(1): 58-62.
- Kaur, G., Kalia, A. and Sodhi, H. 2018. Selenium biofortification of *Pleurotus* species and its effect on yield, phytochemical profiles and protein chemistry of fruiting bodies. *Journal of Food Biochemistry* 42(2): e12467.
- Kayode, R. M. O., Olakulehin, T. F., Adedeji, B. S., Ahmed, O., Aliyu, T. H. and Badmos, A. H. A. 2015. Evaluation of amino acid and fatty acid profiles of commercially cultivated oyster mushroom (*Pleurotus sajor-caju*) grown on gmelina wood waste. *Nigerian Food Journal* 33: 18-21.
- Khandrika, L., Kumar, B., Koul, S., Maroni, P. and Koul, H. K. 2009. Oxidative stress in prostate cancer. *Cancer Letters* 282(2): 125-136.
- Khatun, S., Islam, A., Cakilcioglu, U. and Chatterjee, N. 2012. Research on mushroom as a potential

- source of nutraceuticals: A review on Indian perspective. *American Journal of Experimental Agriculture* 2(1): 47-73.
- Kim, J. H., Kim, S. J., Park, H. R., Choi, J. I., Ju, Y. C., Nam, K. C., ... and Lee, S. C. 2009. The different antioxidant and anticancer activities depending on the color of oyster mushrooms. *Journal of Medicinal Plants Research* 3(12): 1016-1020.
- Koutrotsios, G., Tagkouli, D., Bekiaris, G., Kaliora, A., Tsiaka, T., Tsiantas, K., ... and Zervakis, G. I. 2022. Enhancing the nutritional and functional properties of *Pleurotus citrinopileatus* mushrooms through the exploitation of winery and olive mill wastes. *Food Chemistry* 370: 131022.
- Kumar, K., Mehra, R., Guiné, R. P. F., Lima, M. J., Kumar, N., Kaushik, R., ... and Kumar, H. 2021. Edible mushrooms: A comprehensive review on bioactive compounds with health benefits and processing aspects. *Foods* 10: 2996.
- Lavelli, V., Proserpio, C., Gallotti, F., Laureati, M. and Pagliarini, E. 2018. Circular reuse of bio-resources: The role of *Pleurotus* spp. in the development of functional foods. *Food and Function* 9(3): 1353-1372.
- Lelley, J. 1987. Edible mushrooms as a weapon against starvation. *Mushroom Journal of Tropics* 7: 135-140.
- Li, C. and Xu, S. 2022. Edible mushroom industry in China: Current state and perspectives. *Applied Microbiology and Biotechnology* 106(11): 3949-3955.
- Li, S. and Shah, N. P. 2015. Effects of *Pleurotus eryngii* polysaccharides on bacterial growth, texture properties, proteolytic capacity, and angiotensin-I-converting enzyme-inhibitory activities of fermented milk. *Journal of Dairy Science* 98(5): 2949-2961.
- Li, Y. R., Liu, Q. H., Wang, H. X. and Ng, T. B. 2008. A novel lectin with potent antitumor, mitogenic and HIV-1 reverse transcriptase inhibitory activities from the edible mushroom *Pleurotus citrinopileatus*. *Biochimica et Biophysica Acta* 1780(1): 51-57.
- Llauradó, M. G., Morris-Quevedo, H., Heykers, A., Lanckacker, E., Cappoen, D., Delputte, P., ... and Cos, P. 2021. Differential induction pattern towards classically activated macrophages in response to an immunomodulatory extract from *Pleurotus ostreatus* mycelium. *Journal of Fungi* 7(3): 206.
- Ma, N., Du, H., Ma, G., Yang, W., Han, Y., Hu, Q. and Xiao, H. 2020. Characterization of the immunomodulatory mechanism of a *Pleurotus eryngii* protein by isobaric tags for relative and absolute quantitation proteomics. *Journal of Agricultural and Food Chemistry* 68(46): 13189-13199.
- Maftoun, P., Johari, H., Soltani, M., Malik, R., Othman, N. Z. and El Enshasy, H. A. 2015. The edible mushroom *Pleurotus* spp.: I. Biodiversity and nutritional values. *International Journal of Biotechnology for Wellness Industries* 4(2): 67-83.
- Market Data Forecast. 2022. Latin America mushroom cultivation market research report – Segmentation by type (oyster mushroom, button mushroom, shiitake mushroom, and others) and region - forecast to 2027. Retrieved on February 12, 2023 from Market Data Forecast Website: <https://www.marketdataforecast.com/market-reports/latin-america-mushroom-cultivation-market>
- Marquez-Fernandez, O., Pacheco, L. and Trigos, Á. 2014. Isolation and identification of sterols in a commercial strain of *Pleurotus* sp. *Revista Chapingo Serie Ciencias Forestales y del Ambiente* 20(2): 227-235.
- Milovanovic, I., Stajic, M., Simonić, J., Stanojkovic, T., Knežević, A. and Vukojevic, J. 2014. Antioxidant, antifungal and anticancer activities of Se-enriched *Pleurotus* spp. mycelium extracts. *Archives of Biological Sciences* 66(4): 1379-1388.
- Mizutani, T., Inatomi, S., Inazu, A. and Kawahara, E. 2010. Hypolipidemic effect of *P. eryngii* extract in fat-loaded mice. *Journal of Nutritional Science and Vitaminology* 56(1): 48-53.
- Mohamed E. M. and Farghaly F. A. 2014. Bioactive compounds of fresh and dried *Pleurotus ostreatus* mushroom. *International Journal of Biotechnology for Wellness Industries* 3(1): 4-14.
- Nakahara, D., Nan, C., Mori, K., Hanayama, M., Kikuchi, H., Hirai, S. and Egashira, Y. 2020. Effect of mushroom polysaccharides from *Pleurotus eryngii* on obesity and gut microbiota in mice fed a high-fat diet.

- European Journal of Nutrition 59(7): 3231-3244.
- Ndung'u, S. W., Otieno, C. A., Onyango, C. and Musieba, F. 2015. Nutritional composition, physical qualities and sensory evaluation of wheat bread supplemented with oyster mushroom. American Journal of Food Technology 10(6): 279-88.
- Ng, S. H., Robert, S. D., Wan Ahmad, W. A. and Wan Ishak, W. R. 2017. Incorporation of dietary fibre-rich oyster mushroom (*Pleurotus sajor-caju*) powder improves postprandial glycaemic response by interfering with starch granule structure and starch digestibility of biscuit. Food Chemistry 227:358-368.
- Niazi, A. R. and Ghafoor, A. 2021. Different ways to exploit mushrooms: A review. All Life 14(1): 450-460.
- Nitschke, J., Modick, H., Busch, E., von Rekowski R. W., Altenbach H. J. and Mölleken, H. 2011. A new colorimetric method to quantify β -1,3-1,6-glucans in comparison with total β -1,3-glucans in edible mushrooms. Food Chemistry 127(2): 791-796.
- Nozaki, H., Itonori, S., Sugita, M., Nakamura, K., Ohba, K., Suzuki, A. and Kushi, Y. 2008. Mushroom acidic glycosphingolipid induction of cytokine secretion from murine T cells and proliferation of NK1.1 a/b TCR-double positive cells *in vitro*. Biochemical and Biophysical Research Communications 373(3): 435-439.
- Paulík, S., Švrček, M. J., Ďurove, A., Beníšek, Z. and Húska, M. 1996. The immunomodulatory effect of the soluble fungal glucan (*Pleurotus ostreatus*) on delayed hypersensitivity and phagocytic ability of blood leucocytes in mice. Journal of Veterinary Medicine Series B 43(3): 129-135.
- Perduca, M., Destefanis, L., Bovi, M., Galliano, M., Munari, F., Assfalg, M., ... and Capaldi, S. 2020. Structure and properties of the oyster mushroom (*Pleurotus ostreatus*) lectin. Glycobiology 30(8): 550-562.
- Prabu, M. and Kumuthakalavalli, R. 2016. Antioxidant activity of oyster mushroom (*Pleurotus florida* [Mont.] Singer) and milky mushroom (*Calocybe indica* P&C). International Journal of Current Pharmaceutical Review and Research 8: 48-51.
- Qi, Y., Chen, H., Zhang, M., Wen, Q., Qiu, L. and Shen, J. 2019. Identification and expression analysis of *Pofst3* suggests a role during *Pleurotus ostreatus* primordia formation. Fungal Biology 123(3): 200-208.
- Ragasa, C. 2018. Anticancer compounds from nine commercially grown and wild Philippine mushrooms. Manila Journal of Science 11: 42-57.
- Ragasa, C., Ebajo, J. V., Reyes, R., Brkljača, R. and Urban, S. 2015. Sterols and lipids from *Pleurotus florida*. Der Pharma Chemica 7: 331-336.
- Raman, J., Jang, K. Y., Oh, Y. L., Oh, M., Im, J. H., Lakshmanan, H. and Sabaratnam, V. 2021. Cultivation and nutritional value of prominent *Pleurotus* spp.: An overview. Mycobiology 49(1): 1-14.
- Rana, T., Bera, A. K., Bhattacharya, D., Das, S., Pan, D. and Das, S. K. 2015. Characterization of arsenic-induced cytotoxicity in liver with stress in erythrocytes and its reversibility with *Pleurotus florida* lectin. Toxicology and Industrial Health 31(2): 108-122.
- Rathee, S., Rathee, D., Rathee, D., Kumar, V. and Rathee, P. 2012. Mushrooms as therapeutic agents. Brazilian Journal of Pharmacognosy 22(2): 459-474.
- Ravishankar, S., Pandey, M., Tewari, R. P. and Krishna, V. 2006. Development of sporeless / low sporing strains of *Pleurotus* through mutation. World Journal of Microbiology and Biotechnology 22:1021-1025.
- Ren, D., Jiao, Y., Yang, X., Yuan, L., Guo, J. and Zhao, Y. 2015. Antioxidant and antitumor effects of polysaccharides from the fungus *Pleurotus abalonus*. Chemico-Biological Interactions 237: 166-174.
- Ren, Z., Li, J., Xu, N., Zhang, J., Song, X., Wang, X., ... and Jia, L. 2017. Anti-hyperlipidemic and antioxidant effects of alkali-extractable mycelia polysaccharides by *Pleurotus eryngii* var. *tuolensis*. Carbohydrate Polymers 175: 282-292.
- Rop, O., Mlcek, J. and Jurikova, T. 2009. Beta-glucans in higher fungi and their health effects. Nutrition Reviews 67(11): 624-631.
- Royse, D. J., Baars, J. and Tan, Q. 2017. Current overview of mushroom production in the world. In Diego, C. Z. and Pardo-Giménez, A.

- (eds). Edible and Medicinal Mushrooms - Technology and Applications. United Kingdom: Wiley-Blackwell.
- Saiful Bahri, S. and Wan Rosli, W. I. 2016. Effect of oyster mushroom (*Pleurotus sajor-caju*) addition on the nutritional composition and sensory evaluation of herbal seasoning. *International Food Research Journal* 23(1): 262-268.
- Sánchez, C. 2004. Modern aspects of mushroom culture technology. *Applied Microbiology and Biotechnology* 64(6): 756-762.
- Sánchez, J. E. and Royse, D. J. 2017. World production of *Pleurotus* spp. mushrooms with emphasis on Ibero-American countries. México: El Colegio de la Frontera Sur Tapachula.
- Sande, D., de Oliveira, G., Moura, M., Martins, B., Nogueira, T. and Takahashi, J. 2019. Edible mushrooms as a ubiquitous source of essential fatty acids. *Food Research International* 125: 108524.
- Sarangi, I., Ghosh, D., Kumar Buthia, S., Kumar Mallick, S. and Maiti, T. 2006. Anti-tumor and immunomodulating effects of *Pleurotus ostreatus* mycelia-derived proteoglycans. *International Immunopharmacology* 6(8): 1287-1297.
- Sari, M., Prange, A., Lelley, J. I. and Hambitzer, R. 2017. Screening of beta-glucan contents in commercially cultivated and wild growing mushrooms. *Food Chemistry* 216: 45-51.
- Schneider, I., Kressel, G., Meyer, A., Krings, U., Berger, R. G. and Hahn, A. 2011. Lipid lowering effects of oyster mushroom (*Pleurotus ostreatus*) in humans. *Journal of Functional Foods* 3(1): 17-24.
- Selvamani, S., El-Enshasy, H. A., Dailin, D. J., Malek, R. A., Hanapi, S. Z., Ambehabati, K. K., ... and Molo, N. 2018. Antioxidant compounds of the edible mushroom *Pleurotus ostreatus*. *International Journal of Biotechnology for Wellness Industries* 7: 1-14.
- Selvi, S., Umadevi, P., Murugan, S. and Senapathy, G. 2011. Anticancer potential evoked by *Pleurotus florida* and *Calocybe indica* using T 24 urinary bladder cancer cell line. *African Journal of Biotechnology* 10: 7279-7285.
- Sharma, K. K. and Kuhad, R. C. 2010. Genetic transformation of lignin degrading fungi facilitated by *Agrobacterium tumefaciens*. *BMC Biotechnology* 10: 67.
- Sheng, Y., Zhao, C., Zheng, S., Mei, X., Huang, K., Wang, G. and He, X. 2019. Anti-obesity and hypolipidemic effect of water extract from *Pleurotus citrinopileatus* in C57BL/6J mice. *Food Science and Nutrition* 7(4): 1295-1301.
- Silveira, M. L., Smiderle, F. R., Agostini, F., Pereira, E. M., Bonatti-Chaves, M., Wisbeck, E., ... and Iacomini, M. 2015. Exopolysaccharide produced by *Pleurotus sajor-caju*: Its chemical structure and anti-inflammatory activity. *International Journal of Biological Macromolecules* 75: 90-96.
- Silveira, M. L., Smiderle, F. R., Moraes, C. P., Borato, D. G., Baggio, C. H., Ruthes, A. C., ... and Iacomini, M. 2014. Structural characterization and anti-inflammatory activity of a linear β -D-glucan isolated from *Pleurotus sajor-caju*. *Carbohydrate Polymers* 113: 588-596.
- Sudha, G., Janardhanan, A., Moorthy, A., Chinnasamy, M., Gunasekaran, S., Thimmaraju, A. and Gopalan, J. 2016. Comparative study on the antioxidant activity of methanolic and aqueous extracts from the fruiting bodies of an edible mushroom *Pleurotus djamor*. *Food Science and Biotechnology* 25: 371-377.
- Sun, Y., Zhou, C., Huang, S. and Jiang, C. 2017. Selenium polysaccharide SPMP-2a from *Pleurotus geesteranus* alleviates H₂O₂-induced oxidative damage in HaCaT cells. *Biomed Research International* 2017: 4940384.
- Sunagawa, M. and Magae, Y. 2002. Transformation of the edible mushroom *Pleurotus ostreatus* by particle bombardment. *FEMS Microbiology Letters* 211(2): 143-146.
- Tesfaw, A., Tadesse, A. and Kiros, G. 2015. Optimization of oyster (*Pleurotus ostreatus*) mushroom cultivation using locally available substrates and materials in Debre Berhan, Ethiopia. *Journal of Applied Biology and Biotechnology* 3(1): 15-20.
- Torres-Martínez, B. M., Vargas-Sánchez, R. D., Torrecano-Urrutia, G. R., Esqueda, M., Rodríguez-Carpena, J. G., Fernández-López, J., Perez-Alvarez, J. A. and Sánchez-Escalante, A. 2022. *Pleurotus* genus as a potential ingredient for meat products. *Foods* 11: 779.

- Toyokawa, C., Shobu, M., Tsukamoto, R., Okamura, S., Honda, Y., Kamitsuji, H., ... and Irie, T. 2016. Effects of overexpression of *PKAc* genes on expressions of lignin-modifying enzymes by *Pleurotus ostreatus*. *Bioscience, Biotechnology and Biochemistry* 80(9): 1759-1767.
- Uddin, P. M. M., Islam, M. S., Pervin, R., Dutta, S., Talukder, R. I. and Rahman, M. 2019. Optimization of extraction of antioxidant polysaccharide from *Pleurotus ostreatus* (Jacq.) P. Kumm and its cytotoxic activity against murine lymphoid cancer cell line. *PLoS One* 14: e0209371.
- United Kingdom Conference of the Parties (UKCOP). 2021. COP26 keeps 1.5°C alive and finalises Paris agreement. Retrieved on February 8, 2022 from UKCOP Website: <https://ukcop26.org/cop26-keeps-1-5c-alive-and-finalises-paris-agreement/>
- Wan Rosli, W. I. and Solihah, M. A. 2012. Effect on the addition of *Pleurotus sajor-caju* (PSC) on physical and sensorial properties of beef patty. *International Food Research Journal* 19(3): 993-999.
- Wan Rosli, W. I., Solihah, M. A. and Mohsin, S. S. J. 2011. On the ability of oyster mushroom (*Pleurotus sajor-caju*) conferring changes in proximate composition and sensory evaluation of chicken patty. *International Food Research Journal* 18(4): 1463-1469.
- Wang, D., Sakoda, A. and Suzuki, M. 2001. Biological efficiency and nutritional value of *Pleurotus ostreatus* cultivated on spent beer grain. *Bioresource Technology* 78(3): 293-300.
- Wang, H., Gao, J. and Ng, T. B. 2000. A new lectin with highly potent antihepatoma and antisarcoma activities from the oyster mushroom *Pleurotus ostreatus*. *Biochemical and Biophysical Research Communications* 275(3): 810-816.
- Wang, W., Chen, K., Liu, Q., Johnston, N., Ma, Z., Zhang, F. and Zheng, X. 2014. Suppression of tumor growth by *Pleurotus ferulae* ethanol extract through induction of cell apoptosis, and inhibition of cell proliferation and migration. *PLoS One* 9: e102673.
- Wang, Y., Li, G., Jiao, X., Cheng, X., Abdullah, M., Li, D., ... and Nie, F. 2019. Molecular characterization and overexpression of *mnp6* and *vp3* from *Pleurotus ostreatus* revealed their involvement in biodegradation of cotton stalk lignin. *Biology Open* 8(2): bio036483.
- Wiater, A., Paduch, R., Pleszczyńska, M., Próchniak, K., Choma, A., Kandefler-Szerszeń, M. and Szczodrak, J. 2011. α -(1 \rightarrow 3)-D-glucans from fruiting bodies of selected macromycetes fungi and the biological activity of their carboxymethylated products. *Biotechnology Letters* 33(4):787-795.
- Wiater, A., Paduch, R., Próchniak, K., Pleszczyńska, M., Siwulski, M., Białas, W. and Szczodrak Z. 2015. Assessing biological activity of carboxymethylated derivatives of α -(1 \rightarrow 3)-glucans isolated from fruiting bodies of cultivated *Pleurotus* species. *Żywność Nauka Technologia Jakość* 1(98): 193-206.
- Wisbeck, E., Facchini, J. M., Alves, E. P., Silveira, M. L. L., Gern, R. M. M., Ninow, J. L. and Furlan, S. A. 2017. A polysaccharide fraction extracted from *Pleurotus ostreatus* mycelial biomass inhibit Sarcoma 180 tumor. *Anais da Academia Brasileira de Ciências* 89(3): 2013-2020.
- World Intellectual Property Organization (WIPO). 2020. PATENTSCOPE simple search. Retrieved on July 10, 2020 from WIPO Website: <https://patentscope.wipo.int/search/en/search.jsf>
- Wu, J. Y., Chen, C. H., Chang, W. H., Chung, K. T., Liu, Y. W., Lu, F. J. and Chen, C. H. 2011. Anti-cancer effects of protein extracts from *Calvatia lilacina*, *Pleurotus ostreatus* and *Volvariella volvacea*. *Evidence-Based Complementary and Alternative Medicine* 2011: 982368.
- Xu, C. J., Wang, Y. X., Niu, B. N., Liu, B., Li, Y. B., Wang, X. M. and Lu, S. L. 2014. Isolation and characterization of a novel lectin with mitogenic activity from *Pleurotus ferulae*. *Pakistan Journal of Pharmaceutical Sciences* 27(4): 983-989.
- Xu, N., Ren, Z., Zhang, J., Song, X., Gao, Z., Jing, H., ... and Jia, L. 2017. Antioxidant and anti-hyperlipidemic effects of mycelia zinc polysaccharides by *Pleurotus eryngii* var. *tuoliensis*. *International Journal Biological Macromolecules* 95: 204-214.
- Xu, W., Huang, J. J.-H. and Cheung, P. C. K. 2012. Extract of *Pleurotus pulmonarius* suppresses

- liver cancer development and progression through Inhibition of VEGF-induced PI3K/AKT signaling pathway. *PLoS One* 7(3): e34406.
- Yan, H. and Chang, H. 2012. Antioxidant and antitumor activities of selenium- and zinc-enriched oyster mushroom in mice. *Biological Trace Element Research* 150(1-3): 236-241.
- Yang, Y., Yuan, P., Wie, X., Fu, C., Li, J., Wang, W., ... and Li, J. 2018. Cultivated and wild *Pleurotus ferulae* ethanol extracts inhibit hepatocellular carcinoma cell growth via inducing endoplasmic reticulum stress- and mitochondria-dependent apoptosis. *Scientific Reports* 8: 13984.
- Yin, C., Zheng, L., Zhu, J., Chen, L. and Ma, A. 2015. Enhancing stress tolerance by overexpression of a methionine sulfoxide reductase A (*MsrA*) gene in *Pleurotus ostreatus*. *Applied Microbiology and Biotechnology* 99(7): 3115-3126.
- Yoav, S., Salame, T. M., Feldman, D., Levinson, D., Ioelovich, M., Morag, E., ... and Hadar, Y. 2018. Effects of *cre1* modification in the white-rot fungus *Pleurotus ostreatus* PC9: Altering substrate preference during biological pretreatment. *Biotechnology for Biofuels* 11(1): 1-16.
- Zeb, S. T., Ali, A. B., Ahmad Jafri, S. and Qazi, M. H. 2013. Effect of nicotinic acid (vitamin B₃ or niacin) on the lipid profile of diabetic and non-diabetic rats. *Pakistan Journal of Medical Sciences* 29(5): 1259-1264.
- Zehiroglu, C. and Ozturk Sarikaya, S. B. 2019. The importance of antioxidants and place in today's scientific and technological studies. *Journal of Food Science and Technology* 56(11): 4757-4774.
- Zhang, Q., Yuan, C., Wang, F., Xu, S., Li, Y., Shi, G. and Ding, Z. 2021. Roles of small subunits of laccase (ssPOXA3a/b) in laccase production by *Pleurotus eryngii* var. *ferulae*. *Journal of Agricultural and Food Chemistry* 69: 13113-13124.
- Zisu, B. and Shah, N. P. 2003. Effects of pH, temperature, supplementation with whey protein concentrate, and adjunct cultures on the production of exopolysaccharides by *Streptococcus thermophilus* 1275. *Journal of Dairy Science* 86(11): 3405-3415.